

## SCREENING OF F1 HYBRIDS AND THEIR PARENTS FOR RESISTANCE TO MULTIPLE VIRUSES IN CHILLI (*CAPSICUM ANNUUM* L.)

SHASHIKUMAR. K. T<sup>1</sup> & MADHAVI REDDY. K<sup>2</sup>

<sup>1</sup>Division of Horticulture, University of Agricultural Sciences, G.K.V.K., Bangalore, Karnataka state, India

<sup>2</sup>Division of Vegetable Crops, Indian Institute of Horticultural Research, Bangalore, Karnataka State, India

### ABSTRACT

Among the viruses infecting chili, potato virus Y (PVY), chili veinal mottle virus (CVMV), tobacco etch virus (TEV), pepper vein banding virus (PVBV), pepper veinal mottle virus (PVMV) belonging to potyvirus group and cucumber mosaic virus (CMV) belonging to cucumovirus are economically the most important virus transmitted mechanically, and by aphids in a non persistent manner. In the present investigation, totally 16 parents and their 55 F1 hybrids were screened for four major viruses infecting chili viz., PVY, CVMV, PVBV and CMV. Seedlings were inoculated by mechanical sap inoculation method. Young leaves were observed for symptoms after 2 weeks of inoculation. In parental lines- PMR-57, RHRC-50-1, PMR-76, PMR-37, MI-2, AR-27, AR-28, AR-52, Perennial, Punjab Lal, Punjab Guchedhar, Pant C-1 and Tiwari. Crosses PMR-57 x Perennial, PMR-57 x Tiwari, RHRC-50-1 x Perennial, RHRC-50-1 x Punjab Guchedhar, PMR-69 x Perennial, PMR-69 x Perennial, PMR-69 x Punjab Guchedhar, PMR-69 x Tiwari and AR-28 x Perennial were expressed resistance to at least three viruses tested, and sixteen crosses expressed resistance to at least two viruses. Cross PMR-57 x Punjab Guchedhar expressed resistance to all the four viruses studied.

**KEYWORDS:** Chilli, *Capsicum Annuum*, Virus Resistance, Potyviruses & Cucumber Mosaic Virus

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### INTRODUCTION

About 45 viruses are known to be infected peppers (*Capsicum spp*), of these, 21 viruses are known to be infected in India. Among these, potato virus Y (PVY), chili veinal mottle virus (CVMV), tobacco etch virus (TEV), pepper vein banding virus (PVBV), pepper veinal mottle virus (PVMV) belonging to potyvirus group and cucumber mosaic virus (CMV) belonging to cucumovirus are economically the most important virus transmitted mechanically and by aphids in a non persistent manner (Green and Kim, 1991; Siri Wong *et al.*, 1995; Ravi *et al.*, 1997). Potyviruses and tobamoviruses are destructive in tropical countries (Green and Kim, 1991; Krishna Reddy and Singh, 1995).

The crop losses in Chili vary with variety, virus and region (Koennings and McClure, 1981). Yield loss due to CMV was found to be more than 60 per cent (Joshi and Dubey, 1973; Florini and Zitter, 1987), and 50 per cent due to CVMV (Ong *et al.*, 1980), 40 to 100 per cent due to TMV infection and 70 percent due to TEV infection (Villalon, 1981). The damage caused by viruses depends upon the strains involved, the host cultivars grown and whether the viruses occur singly or in mixed infections.

In India, a few varieties like Pant C-1, Tiwari, Punjab Lal and some other varieties are developed for resistance to viruses in chilies is a common feature in nature, there is a strong need to develop multiple virus

resistant variety/hybrids with desirable fruit type.

The host-virus research has been paying ever increasing attention to seek out plants, resistant to viruses and detecting the occurrence of virus resistance genes. The importance lies in the fact that the chemotherapy and physiotherapy procedures used *in vivo* against viruses are inapplicable in practice, the conventional plant protection methods are of poor efficiency and undesirable from the point of environmental protection. These facts suggest that the control of the virus pathogens of Chile is restricted exclusively to breeding for virus resistant varieties/hybrids.

## **MATERIALS AND METHODS**

### **Screening of Parents and F1 Hybrids for Virus Resistance**

Totally 16 parents and 55 F1 hybrids were screened for four major viruses infecting chili viz., Potato virus Y (PVY), Pepper vein banding virus (PVBV), chili veinal mottle virus (CVMV) and cucumber mosaic virus (CMV) in insect proof screen house.

The parents, which were screened in October 1999 for all four viruses, were again screened. The seeds of all 16 parents and 11 hybrids were sown in seed pans at the first stage of screening. After 10 days, another 11 hybrids were sown; similarly all the hybrids were sown at 10 days interval for convenience. Twenty days old seedlings of each parent and its corresponding hybrids were transplanted to pot with 5 seedlings.

### **Stage of Inoculation**

Seedlings in 3-4 leaf stage were inoculated.

### **Method of Transmission of Virus**

Mechanical sap inoculation: PVY, PVBV, CVMV and CMV, all these viruses can be transmitted mechanically through sap inoculation. Preparation of inoculums of PVY, PVBV, CVMV and CMV: Young leaves showing mosaic symptoms on *Nicotiana glutinosa* were used as a standard source of inoculums of PVY. Young leaves showing mosaic, mottling, vein clearing symptoms on *Nicotiana glutinosa* was used as a standard source of inoculums of PVBV and young leaves showing symptoms of mosaic, mottling on *Datura metal* as standard inoculums of CVMV. Young leaves showing systemic mosaic, and local lesion of *Cucumis sativus* were used as a standard source of inoculums of CMV.

The inoculums were prepared by macerating the leaves in a sterilized pestle and mortar, in the presence of 0.05M phosphate buffer (pH 7.0) at 1:3 (w/v) weights by volume of leaf tissue. After maceration, the pulp was squeezed through sterilized absorbent cotton. The extract was used as standard inoculums for inoculation.

Colette (600 mesh size) was added at the rate of 0.025 g per ml of the standard inoculums. Inoculation was done by dipping the forefinger in the inoculums and simmering gently on the upper surface of the leaves. Then, the inoculated leaves were washed immediately within 5 to 10 minutes, with jet of water from a squeeze bottle, and the plants were kept under observation, till symptoms appeared and advanced, nearly up to 50 days.

## **RESULTS AND DISCUSSIONS**

### **Reaction of Parents and F1 Hybrids Developed to Different Viruses Tested**

#### **Pepper Vein Banding Virus Resistance**

Four testers Perennial, Punjab Cuchedhar, Pant C-1 and Tiwari were found to be susceptible under screen house

condition, whereas, Punjab Lal was found to be tolerant. However, all the testers expressed resistance in the field condition. This indicates the field resistance or vector non-preference. In the present investigation, these lines were not screened against aphid vectors. Out of eleven lines tested, P2, P9 and P10 were found to be tolerant, both under screen house and field condition. The line P7 was found susceptible in the screen house condition and tolerant under field conditions. The tolerance under field conditions may be attributed to natural escape resistance to vectors.

The majority of the crosses expressed susceptibility under screen house and tolerance or resistance under field conditions, which is similar to the house of testers, indicating dominant factors may be responsible. This resistance expression in the field, both in lines and hybrids clearly indicates, resistance is to the insect vector rather than a virus.

Simmonds and Harrison (1956) reported that resistance to PVBV a strain of PVY was due to two unlinked recessive loci. However, this virus is reported as a strain of PVY in 1956 by Simmonds. The present PVBV was isolated from Bangalore and found to be a different virus belonging to potyvirus group. This virus has less than 80 per cent coat protein homology with PVY strains, and therefore, it is a different virus (Joseph and Savitri, 1999). This may be a reason for the resistance behavior to be different from Simmonds and Harrison (1956).

### Potato Virus Y Resistance

All testers used were found to be resistant to PVY, both under screen house and field condition.

Perennial, Punjab Lal and Pant c-1 were earlier reported as resistant to PVY, which are similar to the present investigation (Singh, 1992; Singh and Cheema, 1989; George *et al.*, 1992; Konai and Nariani, 1980). All the lines were found resistant or tolerant, both under screen house and field condition. Out of 55 crosses developed, the majority of the crosses expressed resistance both under screen house and field, whereas, few expressed tolerance. This may be due to the resistance of both the parents. In the course of genetic studies on resistance to PVY, Cook and Anderson (1960), Sharma *et al.*, (1989), Singh and Chenulu (1985) pointed out that, the course of inheritance of resistance was recessive, while the susceptibility of the pepper strains was a dominant hereditary feature. A dominant allele from *Capsicum annuum* line Crillo de Morlos 334, *Pvr* 4 has been conferred dominant resistance to different prototypes (0, 1 and 1,2 of PVY) (Dogimont *et al.*, 1996). Safaris and Marco (1980) reported partial dominance to PVY in *Capsicum*. They indicated that the heterozygous plants are more vigorous and contain lower virus titer than the homozygous plants.

### Chili Veinal Mottle Virus Resistance

Testers Perennial, Punjab Lal and Pant C1 have shown resistance, both under screen house and field condition, whereas, Punjab Guchedhar and Pant C-1 showed tolerance in screen house condition and resistance in field condition. Perennial, Punjab Lal and Pant C-1 and Tiwari were already reported to be resistant to CVMV (Chew and Ong, 1990) Anon, 1993), which is similar to the present result, however, Tiwari was found tolerant in the present investigation.

Susceptible line AR-27, when crossed with resistant testers Perennial, Punjab Lal and Pant C-1, the resulting hybrids were found resistant/tolerant, indicating dominant factors involved in CVMV resistance. Quarantine and Palloix (1996) made similar observations earlier.

Lines PMR-76 and AR-52, which were tolerant to CVMV when crossed with resistant testers, the resulting hybrids were found to be susceptible under screen house conditions and resistant under natural conditions. The susceptibility of F1 generation from tolerant and resistant parents may be due to heterozygous condition. This indicates the

effects of single allele is insufficient even when one allele from each parent is present, resulting in systemic infection (Subramanya, 1982).

### Cucumber Mosaic Virus Resistance

In the present investigation, Perennial found to be susceptible to CMV under screen house conditions, producing mild symptoms and resistant under field condition. This resistance in the field may be due to non-preference of the vector, however, screening with vector was not done in the present investigation. The susceptibility may also be explained by strain variation in CMV (Pinaki Acharya, 1999; Grube *et al.*, 2000). The inconsistency of resistance in Perennial from immune to susceptibility was also reported earlier, by several workers (Singh and Thakur, 1980; Pochard *et al.*, 1983; Singh, 1992).

In the present investigation, Punjab Lal was found to be resistant to CMV under field condition and tolerant under screen house condition. Field resistance to CMV was reported by Singh and Singh (1995), Singh and Cheema (1989), Singh and Kaur (1986), Singh (1992) and Bansal *et al.* (1992). However, they did not report screening under artificial conditions. Pant C-1 and Tiwari were reported to be having field resistance or tolerance to CMV (Singh, 1992; George *et al.*, 1992; Singh and Kaur, 1986). In the present investigation also, Pant C-1 showed tolerance, but Tiwari showed susceptibility to CMV both under field and screen house condition.

The resistant lines PMR-57, RHRC-50-1 and PMR-69 when crossed with field resistant testers, Perennial and Punjab Lal and majority of the resulting hybrids have shown resistance, and a few have shown to CMV indicating dominant factors for resistance.

The best characterized resistance source, Perennial has been described as having monogenic recessive, partially dominant or polygenic recessive resistance, by many workers (Lapidot *et al.*, 1997; Pochard and Dobbeze, 1989; Singh and Thakur, 1997; Grube *et al.* 2000). Similar observations were also seen in the present investigation in various cross combination viz., PMR-69 x Perennial, PMR-37 x Punjab Guchedhar, MI-2 x Tiwari and AR-52 x Punjab Lal.

### Multiple Virus Resistance

Co-segregation of resistance to three potyviruses including pepper mottle virus (PepMoV), tobacco etch virus (TEV) and potato virus Y (PVY) has been observed in pepper (*Capsicum annuum*) for many years (Cook, 1960). A number of *Capsicum* genotypes are resistant to various ranges of potyvirus isolated, are currently considered to be strains of PepMoV, TEV and PVY, and a number of resistant genes have been identified, to which, resistance to more than one virus was attributed (Cook and Anderson, 1960; Dogimont *et al.*, 1996; Kuhn *et al.*, 1989; Kyle and Palloix, 1997; Sowel and Demiski, 1997; Zitter, 1972). In the present investigation also, resistance to poyvirus and cucumovirus was observed in parental lines- PMR-57, RHRC-50-1, PMR-76, PMR-37, MI-2, AR-27, AR-28, AR-52, Perennial, Punjab Lal, Punjab Guchedhar, Pant C-1 and Tiwari. Crosses PMR-57 x Perennial, PMR-57 x Tiwari, RHRC-50-1 x Perennial, RHRC-50-1 x Punjab Guchedhar, PMR-69 x Perennial, PMR-69 x Perennial, PMR-69 x Punjab Guchedhar, PMR-69 x Tiwari and AR-28 x. Perennial were expressed resistance to at least three viruses tested and sixteen crosses expressed resistance to at least two viruses. Cross PMR-57 x Punjab Guchedhar expressed resistance to all the four viruses studied.

## CONCLUSIONS

There is urgent need to develop multiple virus resistant hybrids in chilli. In farmers' fields, multiple virus diseases occur at a time. In the present investigation, we made an attempt to identify the sources of resistance to major viral diseases, by screening sixteen parental lines. Parental lines were crossed in a line x tester mating design to develop 55 F1 hybrids. All the parents and F1 hybrids were screened for resistance to PVY, CVMV, PVBV and CMV viruses. We could identify PMR-57, RHRC-50-1, PMR-76, PMR-37, MI-2, AR-27, AR-28, AR-52, Perennial, Punjab Lal, Punjab Guchedhar, Pant C-1 and Tiwari as resistant lines to potyviruses and cucumber mosaic virus. Crosses PMR-57 x Perennial, PMR-57 x Tiwari, RHRC-50-1 x Perennial, RHRC-50-1 x Punjab Guchedhar, PMR-69 x Perennial, PMR-69 x Perennial, PMR-69 x Punjab Guchedhar, PMR-69 x Tiwari and AR-28 x Perennial expressed resistance to at least three viruses tested and sixteen crosses expressed resistance to at least two viruses. Cross PMR-57 x Punjab Guchedhar found resistant to all the four viruses. Further investigations are necessary to find the nature of gene action.

## REFERENCES

1. Anonymous, 1993. AVRDC Progress Report, Asian Vegetable Research and Development Center, Shanhua, Tainan, Taiwan, ROC. 537p.
2. Bansal, R.D., Aulakh, R.K. and Hundal, J.S., 1992. Reaction of different genotypes of pepper (*Capsicum annuum* L.) to cucumber mosaic virus P 132-137. In: VIIIth meeting on "Genetics and Breeding of Capsicum and Eggplant" 7-10 September 1992, Rome, Italy.
3. Caranta, C. and Palloix, A. 1996. Both common and specific genetic factors involved in polygenic resistance in pepper to several potyviruses. *Theor. Appl. Genet.*, 92(1): 15-20.
4. Chew, B.H. and Ong, C.A. 1980. Genetics and breeding for chilli veinal mottle virus and cucumber mosaic virus resistance in hot pepper p55-58. In: Proc. 3<sup>rd</sup> Int. Conference Plant Protection in the tropics, Vol V. 20-23.
5. Cook, A.A. and Anderson, C.W. 1960. Multiple virus disease in 9 strains of *Capsicum annuum* L. *Phytopathol.* 49: 198-201.
6. Dogimont, C., Palloix, A., Daubeze, A.M., Marchoux, G., Gebre Selassie K, Pochard, E. 1996. Genetic analysis of broad spectrum resistance to potyviruses using doubled haploid lines of pepper (*Capsicum annuum* L.). *Euphytica* 88: 231-239.
7. Florini, D.A. and Zitter, T.A. 1987. Cucumber mosaic virus (CMV) in peppers (*Capsicum annuum* L.) in New York and associated yield losses. *Phytopathol.*, 77: 652 (Abstr.).
8. George, T.E., Anand, N., Deshpande, A.A. and Singh, S.J. 1992. Studies on sources of resistance to potato virus Y (PVY) and cucumber mosaic virus (CMV) in *Capsicum* species. *Indian Hort.*, 6: 621-627.
9. Green, S.K., Kim, J.S. 1991. Characteristics and control of viruses infecting peppers: a literature review. AVRDC Tech Bull. No. 18.
10. Grube, R.C., Zhang, Y., Murphy, J.F., Loaiza-Figueroa, F., Lackney, V.K., Provvidenti, R. and Jahn, M.K. 2000. New source of resistance to cucumber mosaic virus in *Capsicum frutescens*, *Plant Dis.* 84: 885-891.
11. Joseph, J. and Savitri, H.S. 1999. Determination of 3' terminal nucleotide sequence of pepper break vein banding virus RNA and expression of its coat protein *Escherichia coli*. *Arch. Virol.* 144(9): 1679-87.
12. Joshi, R.D. and Dubey, L.N. 1973. Assessment of losses due to CMV in chilli. *Science and Culture*, 39: 521-522.
13. Koennings, S.R. and McClure, M.A. 1981. Interaction of two potyviruses and *Meloidogyne incognata* in chilli pepper. *Phytopathol.*, 71: 404-408.

14. Konai, M. and Nariani, T.K. 1980. Reaction of different chilli varieties and *Capsicum* spp. To mosaic and leaf curl viruses. *Indian Phytopathol.* 33: 155.
15. Krishnareddy, M. and Singh, S.J. 1995. Detection and identification of tobamoviruses infecting pepper: Detection of plant pathogens and their management, AVRDC Tech. Bull. No. 18 pp 91-124.
16. Kuhn, C.W., Nutter, F.W. Jr., Padgett, G.B., 1989. Multiple levels of resistance to tobacco etch virus in pepper. *Phytopathol.* 79: 814-818.
17. Kyle, M.M. and Palliox, A. 1997. Proposed revision of nomenclature of potyvirus to tobacco etch virus in *Capsicum*. *Euphytica* 97: 183-188.
18. Lapidot, M., Paran, I., Ben-Joseph, R., Benttarush, S., Pilowsky, M., Cohen, S. and Shifriss, C. 1997. Tolerance to cucumber mosaic virus in pepper: Development of advanced breeding lines and evaluation of virus level. *Plant Dis.* 81: 185-188.
19. Ong, C.A., Varghese, G. and Poh, T.W. 1980. The effect of chilli veinal mottle virus on yield of chilli (*Capsicum annuum* L.). *Malaysian Agricultural Research and Development Institute (MARDI) Res. Bull.* 8(1): 74-79.
20. Pochard, E. and Doubeze, A.M. 1989. Progressive construction of polygenic resistance to cucumber mosaic virus of a polygenic resistance in pepper. *Seventh Meet. Genet. Breed. Capsicum and Eggplant Newsletter*, 8: 187-192.
21. Pochard, E., Dumas de Vaulx, R. and Florent, A. 1993. Linkage between partial resistance to cmv and susceptibility to TMV in the line Perennial Analysis on androgenetic homozygous lines. *Capsicum Newsletter* 2: 34-35.
22. Ravi, K.S., Joseph, J., Nagaraju, N., Krishnaprasad, S., Reddy, H.R. and Savitri, H.S. 1997. Characterization of pepper vein banding virus from chilli pepper in India. *Plant Dis.* 81: 673-676.
23. Sharma, O.P., Sharma, P.P. and Chawla, S.C. 1989. Inheritance of resistance to potato virus Y in garden pepper (*Capsicum annuum* L.). *Euphytica* 42: 31-33.
24. Shifriss, C and Marco, S., 1980. Partial dominance of resistance to potato virus Y in *Capsicum*. *Plant Dis.* 64: 57-59.
25. Simmonds and Harrison, 1956. The genetics of resistance to pepper vein banding virus. *Genet.* 44: 1281-1289.
26. Singh, 1992. Heterosis studies in chillies (*Capsicum annuum* L.). *Veg. Sci.* 19(2): 161-165.
27. Singh, J.H. and Cheema, D.S. 1989. Present status of tomato and pepper production in India. In: *Tomato and pepper production in the tropics proceedings of the international symposium on integrated management practices, Shanhua, Taiwan: AVRDC* pp 452-471.
28. Singh, S. and Chenulu, V.V. 1986. Studies on resistance to virus diseases in *Capsicum* species III. Inheritance of resistance to Potato virus Y. *Indian Phytopathol.* 38(3): 479-483.
29. Singh, J and Kaur, S., 1986, Present status of hot pepper breeding for multiple disease resistance in Punjab. In VI meeting on genetics and breeding on *Capsicum* and *Eggplant* Zaragoza (Spain), October 21-24.
30. Singh, M.J. and Singh, J., 1995. Sources of resistance to cucumber mosaic virus in chillies. *Plant Dis. Res.* 13(2):184.
31. Singh, J. and Thakur, M.R. 1977. Genetics of resistance to tobacco mosaic virus, cucumber mosaic virus and leaf curl virus in hot pepper (*Capsicum annuum* L.). III EUCARPIA *Capsicum* meeting, Avignon (France), pp 119-126.
32. Singh, S.J. and Thakur, M.R. 1980. Reaction of some hot pepper (*Capsicum annuum* L.) lines to cucumber mosaic virus. *Indian J. Mycol. and Plant Pathol.* 9: 276.
33. Siri Wong, P., Kittipakorn, K. and Ikegami, M. 1995. Characterization of chilli vein banding mottle virus isolated from pepper in Thailand. *Plant Pathol.* 44: 718-727.

34. Sowell, G., and Demski, J.W. 1977. Resistance to plant introductions in pepper to tobacco etch virus. *Plant Dis.* 63: 172-179.
35. Subramanya, R., 1982. Relationship between tolerance and resistance to pepper mottle virus in a cross between *Capsicum annuum* L x *Capsicum frutescens* Jacq. *Euphytica* 31: 461-464.
36. Villalon, B. 1981. Breeding peppers to resist virus diseases. *Plant Dis.*, 65: 557-561.
37. Zitter, T.A. and Cook, A.A., 1972. Inheritance to tolerance to a pepper virus in Florida. *Phytopathol.*, 63: 1211-1212.

## APPENDICES

**Table 1: Reaction of Hybrids to Pepper Vein Banding Virus (PVBV) under Screen House and Field Condition in Chilli**

	Perennial (S,R)		Punjab Lal (T,R)		Punjab Guchedhar (S,R)		Pant C-1 (S,R)		Tiwari (S,R)	
	SH	F	SH	F	SH	F	SH	F	SH	F
PMR-57 (S,T)	R	R	R	R	R	R	R	R	R	R
RHRC-50-1 (T,T)	R	R	R	R	R	R	R	R	R	R
PMR-69(S,S)	R	R	R	R	-	T	R	T	R	T
PMR-76 (S,S)	T	R	T	T	T	T	T	T	R	T
PMR-37 (S,S)	S	T	S	T	T	T	S	T	T	T
MI-2 (T,T)	T	R	T	R	T	R	T	R	T	R
EG-174(S,T)	S	R	S	R	S	R	S	R	S	R
AR-27 (S,T)	S	T	S	R	S	R	R	R	S	R
AR-28 (T,T)	S	R	-	R	R	R	-	R	-	R
AR-52(T,T)	R	R	R	R	R	R	R	R	T	R
D-76 (S,S)	R	R	T	R	T	R	T	R	T	R

R= Resistant, T= Tolerant, S=Susceptible; SH= Screen House, F=Field

**Table 2: Reaction of Hybrids to Potato Virus Y (PVY) under Screen House and Field Condition in Chilli**

	Perennial (R,R)		Punjab Lal (R,R)		Punjab Guchedhar (R,R)		Pant C-1 (R,R)		Tiwari (R,R)	
	SH	F	SH	F	SH	F	SH	F	SH	F
PMR-57 (T,R)	R	R	R	R	R	R	-	R	-	R
RHRC-50-1 (R,T)	R	R	-	R	R	R	-	R	-	R
PMR-69(T,R)	R	R	R	R	-	R	-	R	-	R
PMR-76 (R,R)	-	R	-	R	-	R	-	R	-	R
PMR-37 (R,R)	R	R	R	R	R	R	-	R	-	R
MI-2 (T,R)	R	R	T	R	R	R	R	R	R	R
EG-174(R,R)	R	R	R	R	R	R	S	R	R	R
AR-27 (R,R)	R	R	-	R	-	R	-	R	-	R
AR-28 (R,R)	T	R	-	R	R	R	R	R	T	R
AR-52(R,R)	-	R	-	R	-	R	-	R	T	R
D-76 (R,R)	T	R	-	R	-	R	-	R	-	R

R= Resistant, T= Tolerant, S=Susceptible; SH= Screen House, F=Field

**Table 3: Reaction of Hybrids to Chilli Veinal Mottle Virus (CVMV) under Screen House and Field Condition in Chilli**

	Perennial (R,R)		Punjab Lal (R,R)		Punjab Guchedhar (T,R)		Pant C-1 (R,R)		Tiwarei (T,R)	
	SH	F	SH	F	SH	F	SH	F	SH	F
PMR-57 (T,R)	R	R	R	R	-	R	R	R	R	R
RHRC-50-1 (R,T)	R	R	R	R	R	R	R	R	R	R
PMR-69(T,S)	R	R	R	R	R	R	R	R	R	R
PMR-76 (T,T)	R	R	R	R	-	R	-	T	-	T
PMR-37 (R,T)	S	R	S	R	R	R	S	R	-	R
MI-2 (S,T)	S	R	S	R	S	R	S	R	S	R
EG-174(S,S)	T	R	S	R	S	R	S	R	S	R
AR-27 (S,S)	S	R	T	R	T	R	R	R	S	R
AR-28 (R,T)	S	R	-	R	R	R	T	R	T	R
AR-52(T,T)	-	R	R	R	-	R	-	R	-	R
D-76 (S,S)	-	R	-	R	-	R	-	R	-	R

R= Resistant, T= Tolerant, S=Susceptible; SH= Screen House, F=Field

**Table 4: Reaction of Hybrids to Cucumber Mosaic Virus (CMV) under Screen House and Field Condition in Chilli**

	Perennial (S,R)		Punjab Lal (T,R)		Punjab Guchedhar (S,T)		Pant C-1 (S,T)		Tiwarei (S,S)	
	SH	F	SH	F	SH	F	SH	F	SH	F
PMR-57 (R,R)	R	R	-	R	R	R	-	T	R	R
RHRC-50-1 (R,R)	-	R	-	R	-	R	-	R	-	R
PMR-69(T,S)	R	R	-	R	R	R	R	T	R	R
PMR-76 (R,R)	-	R	-	T	-	T	R	R	-	T
PMR-37 (S,S)	S	R	R	R	-	R	-	R	-	R
MI-2 (S,S)	T	R	R	R	R	R	T	R	R	T
EG-174(S,S)	S	T	T	T	R	R	R	R	R	R
AR-27 (S,T)	T	R	-	R	-	R	-	R	-	R
AR-28 (T,T)	R	R	R	R	-	R	-	T	-	R
AR-52(T,T)	R	R	R	R	-	T	-	R	-	R
D-76 (S,S)	-	R	-	R	-	R	-	R	-	R

R= Resistant, T= Tolerant, S=Susceptible; SH= Screen House, F=Field